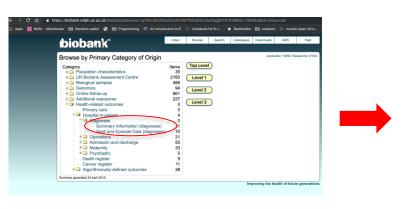
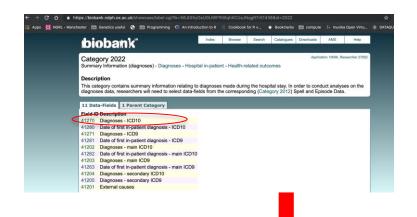
UK Biobank – UoM

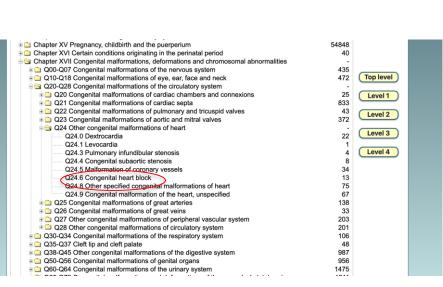
Getting started with hospital episode statistics (HES) data

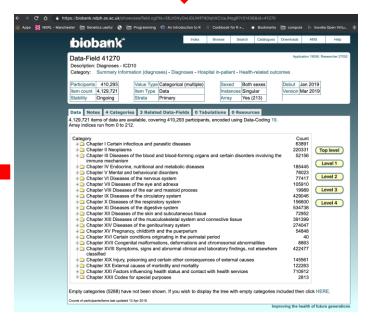
27/06/19

HES – what diagnoses are present?

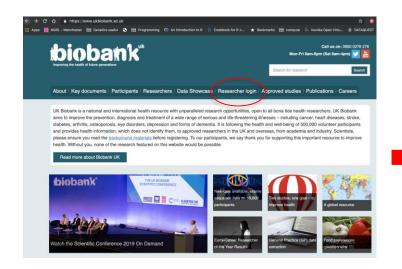


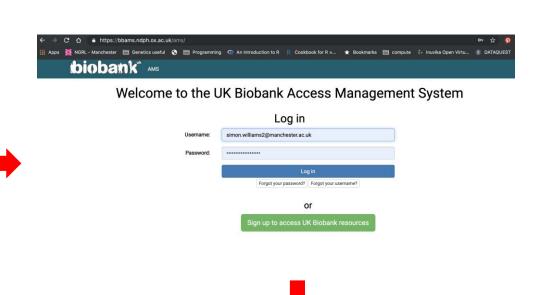


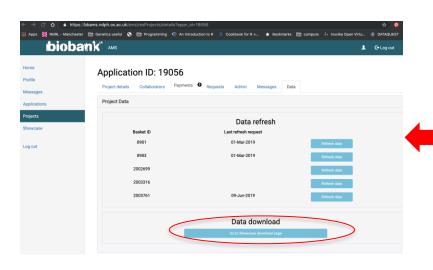


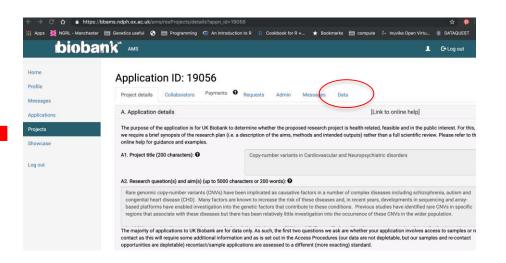


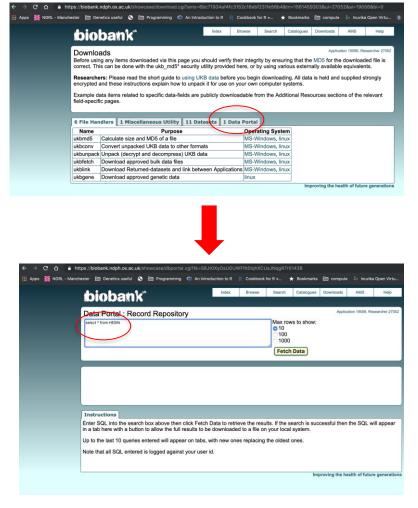
HES – where to access the data once project has been approved











HES tables

HESIN

- main primary hospital diagnoses
- episode/admission dates
- primary ICD9/ICD10/OPCS4 codes

HESIN DIAG10

secondary ICD10 diagnoses

HESIN OPER

secondary OPCS4 codes

HESIN_DIAG9

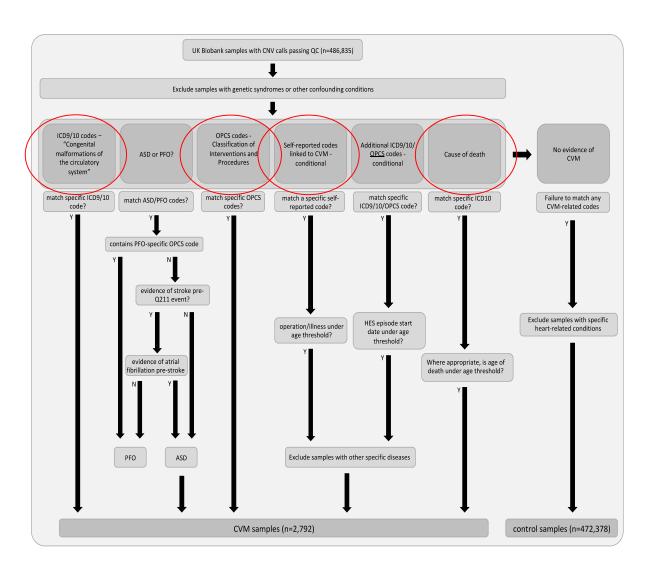
secondary ICD9 codes

Data extracted by SQL queries:

- Query specific code e.g. SELECT eid FROM hesin WHERE diag_icd10 = 'Q256'
- Get everything and export e.g. SELECT * FROM hesin

Rscript to open HES tables and find samples with matching diagnosis - 'Heart block (Q246)'

library(data.table) # load the HES data tables you've exported from UKB SQL page #read hesin table hesin=fread("HESIN.tsv") #read hesin diag10 table (secondary diagnosis table) hesin diag10=fread("HESIN DIAG10.tsv") #read hesin oper table (secondary operations table) hesin oper=fread("HESIN OPER.tsv") # extract samples that might be of interest based on ICD10 codes #read list of ICD codes we're interested in - here jut a file with 'Q246' under the 'ICD10' header. codes<-read.csv("heart block phenotype codes", header=TRUE) #subset these samples from HESIN table hit1<-subset(hesin, hesin\$diag icd10 %in% codes\$ICD10) #then HESIN DIAG10 table - this table contains the secondary diagnosis (only ICD10) hit2<-subset(hesin diag10, hesin diag10\$\diag icd10 \%in\% codes\$ICD10) #you might also search for related ICD9/operation codes in the same way #extract the eids eid<-c(hit1\$eid,hit2\$eid) #get unique eids heart block samples<-unique(eid)



Getting started genetic data

What is available?

Arrays

- 488,766 individuals
- 820,967 SNP and indel markers included
- 2 arrays
 - Affymetrix Axiom UK BiLEVE array (~50,000)
 - Affymetrix Axiom UK Biobank array (~450,000)
- Description of the files available:

http://www.ukbiobank.ac.uk/wp-content/uploads/2017/07/ukb genetic file description.txt

Versions

- V1 1st release ~150,000 samples
- V2 full release ~500,000 samples
- V3 for imputation files only due to an error in the initial V2 imputation release these files were re-processed and re-released

CSF3 central dataset

To access you must be added to a group with correct permissions (dataset-ukbiobank-full group)

Email the research IT team with confirmation of approved UKB data access

module load tools/env/ukbiobank-full-release-2018

Sets a number of environment variables:

UKBB FILELIST

/mnt/data-sets/ukbiobank/full-release/filelist.2018.txt – list paths to files

More info:

http://ri.itservices.manchester.ac.uk/csf-apps/software/applications/ukbiobank/

Calls

- The genotype calls are in binary PLINK format (.bed, .bim, .fam) see
 https://www.cog-genomics.org/plink/1.9/formats for details of the formats.
- The BIM file determines the order of markers in the calls and all of the other genotype data sets. The SNP_id is the rsid where it is available or the Affymetrix_SNP_id otherwise.
- The positions are GRCh37 coordinates.
- The **FAM** file determines the order of samples in the calls and all of the other genotype data sets. The FAM file includes 'Batch' in the Phenotype field (6th column). this file is project specific the eids are different between projects

Imputed data

The imputed genotype calls are in BGEN v1.2 format (.bgen, .sample, .bgi)

Files

•	Calls BED	ukb_cal_chrN_v2.bed
•	Calls BIM	ukb_snp_chrN_v2.bim
•	Calls FAM	ukbA_cal_v2_sP.fam Project-specific – download this yourself "ukbgene evc -c1 –m" from linux command line
•	Marker-QC	ukb_snp_qc.txt
•	Sample-QC	ukb_sqc_v2.txt
•	Relatedness	ukbA_rel_sP.txt
•	Imputation BGEN	ukb_imp_chrN_v3.bgen
•	Imputation BGI	ukb_bgi_chrN_v3.bgi
•	Imputation MAF+info	ukb_mfi_chrN_v3.txt
•	Imputation sample	ukbA_imp_autosome_v3_sP.sample
•	Haplotypes BGEN	ukb_hap_chrN_v3.bgen
•	Haplotypes BGI	ukb_hbg_chrN_v3.bgi
•	HLA Imputation	ukb_hla_v2.txt
•	Intensity	ukb_int_chrN_v2.bin
•	Confidences	ukb_con_chrN_v2.txt
•	CNV log2r	ukb_l2r_chrN_v2.txt
•	CNV baf	ukb_baf_chrN_v2.txt
•	SNP-posterior	ukb_snp_posterior_chrN.bin
•	Batch	ukb_snp_posterior.batch

Whole Exome Sequencing

- 1st 50,000 released
- 39Mbp exome
- 75bp paired end reads
- Illumina NovaSeg 6000
- Mapped to GRCh38 reference
- Variant called through two pipelines:
 - 'FE' 'Functional Equivalent' pipeline (GATK)
 - 'SPB' Regeneron's Seal Point Balinese pipeline
- PLINK format release of all samples together
- Individual gVCFs can also be downloaded
- Download using 'ukbgene' utility

Category 170

Exome sequences - Genomics

Description

The first tranche of UKBiobank whole exome sequencing (WES) is now available for ~50,000 UK Biobank participants.

To ensure equality of access the individual level data is currently embargoed to allow all researchers an opportunity to download the PLINK formatted data. The VCF files will be released by early-April followed by the CRAM files. Researchers who already have access to UK Biobank genetic data do NOT have to submit new baskets to request exome data - this will be done for them automatically by the Access team.

This sample set prioritizes individuals with whole body MRI imaging data, enhanced baseline measurements, hospital episode statistics (HES), and/or linked primary care records. Additionally, one disease area was selected for enrichment: individuals with admission to hospital with a primary diagnosis of asthma (ICD10 J45 or J46). The sequenced set includes 194 parent-offspring pairs, 613 full-sibling pairs, including 26 trios, 1 monozygotic twin pair and 195 second degree genetically determined relationships.

Exomes were captured with the IDT xGen Exome Research Panel v1.0 including supplemental probes. The basic design targets 39Mbp of the human genome (19,396 genes). Multiplexed samples were sequenced with dual-indexed 75x75pp paired-end reads on the Illumina NovaSeq 6000 platform using S2 flow cells. In each sample and among targeted bases, coverage exceeds 20X at 94.6% of sites on average. Complete sequencing protocols are described in detail by the summary manuscript (add link when available). This manuscript also fully describes the "SPB" primary and secondary analysis pipeline that converts raw sequencing data to a quality-controlled set of population variation. The SPB pipeline first converted all raw sequencing data to FASTQs according to Illumina NovaSeq best practices and aligned those reads to the GRCh38 reference genome with BWA-mem to generate a CRAM file for each sample. After read-duplicate marking, SNVs and indels were called for with WeCall (GenomicsPLC), generating a gVCF per sample. These gVCFs were joint genotyped using GLnexus (https://www.biorxiv.org/content/10.1101/572347v1) to create a single, unfiltered project-level VCF (pVCF). Genotype depth filters (SNV DP>7, indel DP>10) were applied prior to variant site filters requiring at least one variant genotype passing an allele balance filter (heterozygous SNV AB>0.15, heterozygous indel<0.20), resulting in a second 'filtered' pVCF. A total of 4,735,722 variants are identified within targeted regions, with 9,693,536 variants identified across all covered bases including 100bp regions flanking the capture targets.

To maximize data utility and ease of use, an additional "Functionally Equivalent" (FE) pVCF was generated from FASTQs, following the primary analysis protocol described in the 2018 manuscript (PMID: 30279509) and then subject to GATK 3.0 variant calling and hard filtering of variants with inbreeding coefficient<-0.03 or without at least one variant genotype of DP≥10, GQ≥20 and, if heterozygous, AB≥0.20.

```
Field ID Description

23170 Population-level SPB variants, PLINK format ‡
23160 Population-level Fe variants, PLINK format ‡
23171 Exome SPB variant call files (VCFs) ‡
23172 Exome SPB CRAM files ‡
23173 Exome SPB CRAM files ‡
23174 Exome SPB CRAM indices ‡
23174 Exome SPB CRAM indices ‡
23161 Exome FE variant call files (VCFs) ‡
23162 Exome FE variant calls indices ‡
23163 Exome FE CRAM files ‡
23164 Exome FE CRAM indices ‡
23165 Exome FE CRAM indices ‡
23166 Exome FE CRAM indices ‡
23167 Exome FE CRAM indices ‡
23168 Exome FE CRAM indices ‡
```

Variant call data for 50,000 exomes (sample level VCF files) ~10TB

CRAM files also available ~150TB

Capacity for downloading centrally.....

Initial comparison of arrays and exomes...

Mike Weedon (twitter) - analysis of 3000 QC'd exome SNPS reveals most SNPs on array with MAF <0.005% are FPs

